# ANTIDIABETIC ACTIVITY OF *MOLLUGO NUDICAULIS* AGAINST ALLOXAN INDUCED DIABETIC RATS

# T.Sindhu<sup>1</sup>, S.Rajamanikandan<sup>1</sup>, P.Ragavendran<sup>2</sup>, D.Sophia<sup>2</sup>, P.Meenakshi<sup>3</sup>, D.Durga priya<sup>1</sup> and V.K.Gopalakrishnan<sup>1,2,3\*</sup>

<sup>1</sup>Department of Bioinformatics, Karpagam University, Coimbatore- 641 021 (T.N.), INDIA,

<sup>2</sup>Department of Biochemistry, Karpagam University, Coimbatore- 641 021 (T.N.), INDIA,

<sup>3</sup>Department of Biochemistry, Karpagam Arts and Science College, Coimbatore- 641 021 (T.N.), INDIA,

**ABSTRACT:** The present study was designed to investigate the in alloxan (120 mg/kg b.wt) induced diabetic rats. The ethanolic extract of the whole plant of *Mollugo nudicaulis* (200mg/kg) administered orally to the diabetic rats for 21 days, produced significant decrease in the level of blood glucose, cholesterol, triglycerides, low density lipoprotein (LDL), lipid peroxidation, liver glycogen, serum creatinine, urea, uric acid and liver marker enzymes such as AST, ALT, ALP. It also produced significant increase in High density lipoprotein (HDL), Superoxide dismutase (SOD), Catalase (CAT), Glutathione peroxidase (GPx), Glutathione-S-Transferase (GST), Reduced glutathione (GSH), Vitamin C, which clearly show the antioxidant property of extract. The effect of the ethanolic the extract of *Mollugo nudicaulis* was compared with the standard drug Glibenclamide (1.25mg/kg b.wt).

Keywords: Mollugo nudicaulis, Alloxan, Antidiabetic, Glibenclamide, Antioxidants.

# INTRODUCTION

Diabetes mellitus is a syndrome, which is characterized by hyperglycemia, lipo protein abnormalities, raised basal metabolic rate, defect in enzymes and high oxidative stress induced damage to pancreatic beta cells (Sharma *et. al.*, 2010). Type II diabetes mellitus developed by metabolic abnormalities such as impaired insulin secretion, increased hepatic glucose production and decreased insulin-stimulate of glucose uptake in peripheral (Kakadiya *et. al.*, 2010). Several distinct types of diabetes mellitus exist and are caused by a complex interaction of genetics and environmental factors (Chitra *et. al.*, 2010). The prevalence of diabetes for all age groups world wide was estimated to be 4.4% in 2030 (Chauhan *et. al.*, 2010). In india the statistical projection that the number of diabetes will raise to 57 million in 2025, which was only 15 million in 1995 (Sikarwar and Patil, 2010).

From ancient period, people are using medicinal plants for the treatment of diabetes and WHO estimates that 80% of the populations presently use herbal medicine for primary health care (Atmakuri and Dathi, 2010). Anti diabetic plants has the ability to restore the function of damaged pancreatic tissue by increasing the insulin or inhibiting the intestinal absorption of glucose (Malviya *et.al.*, 2010). Administration of appropriate antioxidants from plant source could prevent or retard the diabetic complications to some extent (Muthulingam, 2010).

International Journal of Applied Biology and Pharmaceutical Technology Page:511 Available online at <u>www.ijabpt.com</u>



*Mollugo nudicaulis* is a wild medicinal herb, used by traditional practitioners to cure whooping cough, jaundice and wound healing (Nagesh, 2008). It is useful mainly in preventing small ring worms in stomach, to control diabetes and to increase the life energy of blood cells (Rao, 2009). *Mollugo nudicaulis* is also used to treat wounds, cold, cough, fever, and body pain (Ragupathy *et.al.*, 2008). The present study was investigated to evaluate the anti-diabetic activity of *Mollugo nudicaulis* against alloxan induced diabetic rats.

# MATERIALS AND METHODS

#### Plant material and extraction

The whole plant of *Mollugo nudicaulis* was collected from Pollachi, Coimbatore district and authenticated by Dr.G.V.S. Murthy, Botanical survey of India, TNAU campus, Coimbatore, Tamilnadu, India. The voucher specimen was deposited to the Botanical Survey of India (No.BSI/SRC/5/23/10-11/Tech.420). The plant material was washed with water, dried in shade and pulverized in grinder-mixer to obtain a coarse powder. About 100 gms of the powder were continuously extracted with 500 ml of ethanol using soxhelt apparatus upto 48 hours. The residue was filtered and concentrated in rotatory evaporator at 40°C under reduced pressure until the solvent has been removed to give an extract. The obtained extract was stored in air tight container at 4°C.

# **Experimental animals**

The female albino rats (150-180 gms) were procured from Karpagam University, Coimbatore, India. The animals were housed in group of 6 rats per cage and maintained under standard laboratory condition at  $24\pm$  2°C in a light controlled room (12 h dark/12 h light) and were provided commercial pellet diet, purified drinking water *ad libitum*. The study was approved by Institutional Animal Ethical Committee (IAEC) constituted for the purpose of CPCSEA. Govt of India.

# **Experimental design**

The rats were divided into five groups of 6 rats each.Group I– Normal control ratsGroup II– Alloxan treated control rats (120 mg/kg b.wt)Group III– Alloxan + ethanolic extract of Mollugo nudicaulis (200mg/kg b.wt)Group IV– Normal rats + ethanolic extract of Mollugo nudicaulis (200mg/kg b.wt)Group V – Alloxan + standard drug Glibenclamide (1.25mg/kg b.wt)

# Induction of diabetes

Type II diabetes was induced to female albino rats by a single intraperitoneal (i.p) injection of alloxan monohydrate (120 mg/kg b.wt) in sterile normal saline (0.9%). The diabetic state was determined after 3 days of alloxination by high blood glucose level and loss of body weight. At the end of 21 days treatment, blood glucose level was estimated by one touch glucometer and rats were sacrificed under chloroform anesthesia. Blood was collected and centrifuged at 3000 rpm for 20 minutes to separate the serum. Liver was removed and washed with ice cold normal saline (0.9%) to remove the blood. About 1 g of the liver tissue was homogenized using 0.1 M Tris – Hcl buffer at pH 4.7 and the supernatant was separated. Serum and the supernatant were used to analyze the biological parameters within 24 hours of sacrifice.

International Journal of Applied Biology and Pharmaceutical Technology Page:512 Available online at <u>www.ijabpt.com</u>



#### **Biochemical estimations:**

Determination of Non-Protein nitrogenous compounds

Non-protein nitrogenous compounds such as urea (Natelson *et.al*, 1951), uric acid (Caraway, 1963), creatinine (Brod, 1948) were assayed in serum.

Determination of Lipid profile

Lipid profile such as cholesterol, triglycerides, HDL and LDL were estimated in serum by diagnostic kit method.

Determination of Transaminase

ALP, AST and ALT (King, 1965) were estimated in serum.

Determination of protein

Protein (Lowry et.al, 1951) was estimated in the homogenate of liver.

Estimation of tissue lipid peroxidation

Lipid peroxidation was estimated (Hogberg *et.al*, 1974) and it was calculated on the basis of the molar extinction coefficient of malondialdehyde (MDA) and expressed in terms nanomolar of MDA/mg protein.

Antioxidant assays

The enzymic antioxidants such as superoxide dismutase (Misra and Fridovich, 1972) catalase (Lueck, 1965) glutathione peroxidase (Rotruck *et.al*, 1973) glutathione-S- transferase (Habig *et.al*, 1974) and non enzymatic antioxidants activity such as vitamin C (Omaye *et.al*, 1979) glutathione (Moran *et.al*, 1979) were evaluated in liver tissue homogenate.

#### Statistical analysis

The results were expressed as mean  $\pm$  SD. The significant of the data was calculated in SPSS software package (10.0) followed by one way ANOVA and were considered statistically significant when p<0.05.

#### RESULTS

Table 1 shows the blood glucose level of diabetic and control rats. The diabetic rats treated with *Mollugo nudicaulis* and glibenclamide which was significantly reduced the level of blood glucose compared with Group II diabetic rats.

Table 2 summarizes the concentration of serum creatinine, urea and uric acid were found to be significantly increased in Group II diabetic control rats when compared to Group I normal control rats. When *Mollugo nudicaulis* was administered to Group III rats, the above parameters were reversed.

# Table 1. Effect of ethanolic extract of Mollugo nudicaulis on the levels of glucose in serum of control and experimental animals

Groups	Glucose
Normal control	110.57±0.87ª
Diabetic control	300.78±0.71°
	$197.74 \pm 0.78^{bc}$
Diabetic control + PE (200mg/kg)	154.51±1.07 <sup>b</sup>
Diabetic control + Glibenclamide	$120.574 \pm 1.42^{a}$
PE alone (200mg/kg)	

Values are expressed as mean  $\pm$  SD for six animals. Values not sharing common superscript letters (a-e) differ significantly at p < 0.05 (DMRT).

Units: Glucose: mg/dl.

International Journal of Applied Biology and Pharmaceutical Technology Page:513 Available online at <u>www.ijabpt.com</u>



Groups	Urea(mg/dl)	Uric acid(mg/dl)	Creatinine(mg/dl)
Normal control	29.85±0.57ª	$8.54{\pm}0.48^{a}$	1.27±0.27ª
Diabetic control	60.24±0.57°	19.2±0.24 <sup>e</sup>	$2.87{\pm}0.48^{d}$
Diabetic Control + PE (200mg/kg)	47.41±1.21 <sup>b</sup>	$10.4{\pm}0.21^{d}$	1.88±0.61°
Diabetic Control + Glibenclamide	$37 \pm 0.85^{b}$	9.0±0.273°	$1.54{\pm}0.87^{b}$
PE alone (200mg/kg)	33.12±0.84 <sup>a</sup>	$8.78 {\pm} 0.27^{b}$	$1.34{\pm}0.47^{a}$

 Table 2. Effect of ethanolic extract of Mollugo nudicaulis on serum

 biochemical parameters in control and experimental animals

Values are expressed as mean  $\pm$  SD for six animals. Values not sharing common superscript letters (a-e) differ significantly at p < 0.05 (DMRT).

Table 3 shows that there was a significant increase in the level of cholesterol, triglycerides, LDL, and decrease in the level of HDL Cholesterol in Group II diabetic control rats when compared to Group I normal control rats. The above parameters were significantly reversed on Group III diabetic rats treated with *Mollugo nudicaulis*.

 Table 3. Effect of ethanolic extract of Mollugo nudicaulis on lipid profile in control and experimental animals

Groups	Cholesterol (mg/dl)	Triglycerides (mg/dl)	HDL(mg/dl)	LDL(mg/dl)
Normal control	65.18±1.27 <sup>e</sup>	85.27±0.89°	$22.87 \pm 0.58^{d}$	$16.57 \pm 0.78^{d}$
Diabetic control	$28.42{\pm}1.47^{a}$	$32.48{\pm}1.57^{a}$	$12.67 \pm 1.15^{a}$	$9.12{\pm}0.87^{a}$
Diabetic Control + PE (200mg/kg)	$41.48 \pm 1.57^{b}$	$52.48 {\pm} 0.89^{b}$	$14.37{\pm}0.87^{b}$	$11.48 {\pm} 0.99^{b}$
Diabetic Control + Glibenclamide	$51.89{\pm}0.85^{\circ}$ $63.47{\pm}0.98^{d}$	$68.49 \pm 0.89^{\circ}$ $81.29 \pm 0.95^{\circ}$	$17.59 \pm 0.89^{\circ}$ $22.98 \pm 0.78^{d}$	13.48±1.07° 16.98±0.89 <sup>d</sup>
PE alone (200mg/kg)				

Values are expressed as mean  $\pm$  SD for six animals. Values not sharing common superscript letters (a-e) differ significantly at p < 0.05 (DMRT).

Table 4 shows the level of liver marker enzymes such as ALP, AST and ALT which were found to be significantly increased in Group II rats. On the other hand, the levels of enzymes were observed to have decreased markedly in the diabetic animals treated with *Mollugo nudicaulis* and glibenclamide.

Table 5 and Table 6 shows the level of liver glycogen and total protein which was found to be decreased significantly in Group II diabetic rats, whereas the levels were in Group III rats administered with the ethanolic extract of *Mollugo nudicaulis*.

 

 Table 4. Effect of ethanolic extract of Mollugo nudicaulis on liver marker enzymes in serum of control and experimental animals

Groups	ALP	AST	ALT
Normal control	108.45±1.24ª	16.87±1.41ª	22.47±0.87ª
Diabetic control	248.49±2.78°	$30.47 {\pm} 0.57^{\circ}$	$41.48{\pm}0.98^{\text{d}}$
	$150.87 \pm 1.47^{b}$	22.75±0.48 <sup>b</sup>	$29.47{\pm}0.87^{ab}$
Diabetic Control + PE (200mg/kg)	121.5±1.89 <sup>ab</sup>	20.47±0.19 <sup>b</sup>	25.12±1.25 <sup>ab</sup>
Diabetic Control + Glibenclamide	$110.48 \pm 1.89^{a}$	$18.51 \pm 0.76^{a}$	$24.87{\pm}0.89^{a}$
PE alone (200mg/kg)			

Values are expressed as mean  $\pm$  SD for six animals. Values not sharing common superscript letters (a-e) differ significantly at p < 0.05 (DMRT). Units: AST, ALT, - µmoles of pyruvate liberated/l; ALP - µmoles of phenol liberated/l

International Journal of Applied Biology and Pharmaceutical Technology Page:514 Available online at <u>www.ijabpt.com</u>



# Table 5. Effect of ethanolic extract of *Mollugo nudicaulis* on the levels of glycogen in liver of control and experimental animals

Groups	Glycogen
Normal control	19.45±0.89 <sup>b</sup>
Diabetic control	$5.02{\pm}0.45^{a}$
	$12.07 \pm 0.12^{ab}$
Diabetic control + PE (200mg/kg)	16.48±0.52 <sup>b</sup>
Diabetic control + Glibenclamide	$18.48 \pm 0.78^{b}$
PE alone (200mg/kg)	

Values are expressed as mean  $\pm$  SD for six animals. Values not sharing common superscript letters (a-e) differ significantly at p < 0.05 (DMRT).Units: Glycogen: mg/g tissue.

#### Table 6. Effect of ethanolic extract of Mollugo nudicaulis on protein level of liver in control and experimental animals

Groups	Protein(mg/dl)
Normal control	158.89±0.91°
Diabetic control	$69.71 \pm 0.87^{a}$
Diabetic Control + PE (200mg/kg)	$127.29 \pm 1.78^{b}$
Diabetic Control + Glibenclamide	136.87±2.78 <sup>b</sup> 152.17±0.17 <sup>c</sup>
PE alone (200mg/kg)	

Values are expressed as mean  $\pm$  SD for six animals. Values not sharing common superscript letters (a-e) differ significantly at p < 0.05 (DMRT).

Table 7 shows the concentration of LPO of normal and experimental rats. It was found to be significant elevation in Group II diabetic rats. A significant decrease was observed in Group III rats treated with Mollugo nudicaulis

Table 8 shows the non enzymatic antioxidants such as GSH and Vitamin C which was found to be significantly decreased in Group II diabetic rats. In contrast, Group III animals treated with Mollugo nudicaulis showed significant increase in both GSH and Vitamin C levels

#### Table 7. Effect of ethanolic extract of *Mollugo nudicaulis* on the levels of lipid peroxide liver of control and experimental animals

Groups	LPO	
Normal control	$2.82{\pm}0.12^{ab}$	
Diabetic control	4.14±0.25°	
	2.76±0.01ª	
Diabetic Control + PE (200mg/kg)	$2.81\pm0.21^{ab}$	
Diabetic Control + Glibenclamide	$2.86\pm0.08^{b}$	
PE alone (200mg/kg)		

Values are expressed as mean  $\pm$  SD for six animals. Values not sharing common superscript letters (a-e) differ significantly at p < 0.05 (DMRT). Units: LPO - nM/mg protein.

International Journal of Applied Biology and Pharmaceutical Technology Page: 515 Available online at www.ijabpt.com

# <u>IJABPT</u>

#### ISSN 0976-4550

enzymic antioxidants in liver of control and experimental animals				
Groups	SOD	CAT	GPx	GST
Normal control	$6.81 \pm 0.27^{d}$	$1.83 \pm 0.48^{d}$	4.30±0.57 <sup>e</sup>	100.32±0.89°
Diabetic control	$3.47{\pm}0.57^{a}$	$0.53{\pm}0.57^{a}$	$1.02{\pm}0.27^{a}$	$60.67{\pm}0.81^{a}$
Diabetic Control + PE (200mg/kg)	$5.17 \pm 0.78^{b}$	$1.28 \pm 0.37^{b}$	$2.83{\pm}0.42^{b}$	$87.51 {\pm} 0.97^{b}$
Diabetic Control + Glibenclamide		$1.56\pm0.89^{\circ}$ $1.81\pm0.27^{\circ}$	$3.14\pm0.28^{\circ}$	$92.47 \pm 0.87^{b}$ $97.91 \pm 0.78^{c}$
$\mathbf{DE}$ along $(200 \text{mg/l} \text{s})$	$0.12 \pm 0.58^{\circ}$	$1.81\pm0.27^{\circ}$	4.14±0.5/*	$9/.91\pm0.78^{\circ}$

 Table 8. Effect of ethanolic extract of Mollugo nudicaulis on the activities of enzymic antioxidants in liver of control and experimental animals

#### PE alone (200mg/kg)

Values are expressed as mean  $\pm$  SD for six animals. Values not sharing common superscript letters (a-e) differ significantly at p < 0.05 (DMRT).

**Units** -SOD - inhibition of 50% nitri formation/min/mg protein; CAT -  $\mu$ mol of H<sub>2</sub>O<sub>2</sub> consumed/min/mg protein; GPx -  $\mu$ mol of glutathione oxidized/min/mg protein;GST-  $\mu$ mole of glutathione utilized/min/ mg protein.

Table 9 shows the concentration of LPO of normal and experimental rats. It was found to be significant elevation in Group II diabetic rats. A significant decrease was observed in Group III rats treated with *Mollugo nudicaulis*.

# Table 9. Effect of ethanolic extract of Mollugo nudicaulis on the activities of non-enzymic antioxidants in liver of control and experimental animals

Groups	GSH	Vit C
Normal control	12.48±0.15°	$1.48 \pm 0.12^{d}$
Diabetic control	$5.57{\pm}0.48^{a}$	$0.84{\pm}0.47^{a}$
	$7.08 \pm 0.87^{b}$	1.15±0.87 <sup>bc</sup>
Diabetic Control + PE (200mg/kg)	8.97±1.58 <sup>b</sup>	1.28±0.94°
Diabetic Control + Glibenclamide	11.41±0.84°	$1.45\pm0.89^{d}$
PE alone (200mg/kg)		

Values are expressed as mean  $\pm$  SD for six animals. Values not sharing common superscript letters (a-e) differ significantly at p < 0.05 (DMRT). Units: Units : GSH, vitamin C

The result shows that there was no significant difference between Group III diabetic rats administered with *Mollugo nudicaulis* and Group V diabetic rats treated with the standard drug Glibenclamide. The level of the above biochemical parameters in Group IV normal rats treated with *Mollugo nudicaulis* shows that there is no significant difference when compared to Group I normal rats.

# DISCUSSION

The result of the present study confirms that the administration of the ethanolic extract of *Mollugo nudicaulis* possesses antidiabetic activity against alloxan induced diabetic rats. The anti-diabetic effect of *Mollugo nudicaulis* may be due to increased release of insulin from the existing  $\beta$ -cells of pancreas similar to that observed after glibenclamide administration.

Glibenclamide is a known sulfonylurea drug which is effective in moderate diabetic state, and ineffective in severe diabetic animals where pancreatic  $\beta$ -cells are almost totally destroyed (Tatiya *et.al.* 2010).

International Journal of Applied Biology and Pharmaceutical Technology Page:516 Available online at <u>www.ijabpt.com</u>

<u>IJABPT</u>

ISSN 0976-4550

Diabetic dyslipidaemia is marked by alloxan in diabetic rats by elevated triglycerides, cholesterol, low density lipoprotein (LDL) and decreased high density lipoprotein (HDL), constitutes an important cardiovascular risk factors (Okoli *et.al.*, 2010) and were reversed in *Mollugo nudicaulis* treated rats. The elevation of liver biomarker enzymes, such as AST, ALT and ALP in diabetic rats indicates the hepatic damage (Rathod *et.al.*, 2009). Rats treated with *Mollugo nudicaulis*, showed its ability to restore the normal functional status of the damaged liver.

As seen in the present study the level of renal function markers such as serum creatinine, urea and uric acid which was increased in diabetic rats indicates the dysfunction of kidney (Kakadiya, 2010), were inhibited by the treatment with *Mollugo nudicaulis* revealing that it exhibits potent antidiabetic activity. Administration of alloxan causes decrease in glycogen content due to enhanced glycogenolysis, which is due to insulin deficiency. So the normal capacity of the liver to synthesize glycogen is impaired (Dheer, 2010). A significant increase in the liver glycogen by administration of *Mollugo nudicaulis*, may be due to an increase level of insulin by it.

Lipid peroxidation was induced by glucose through activation of lipoxygenase enzymes (Kaimal, 2010). Free radical induced lipid peroxidation has gained much importance because of its involvement in several pathologies such as ageing, wound healing, oxygen toxicity, liver disorders, inflammation, etc (Gupta, 2010). The increased level of lipid peroxidation induced tissue injury was observed in diabetic rats. The free radical scavenging activity of Mollugo *nudicaulis inhibited* the lipid peroxidation.

The antioxidant enzymes, such as SOD, CAT, GST and GPx constitute a mutually supportive team of defense against ROS. GST eliminating toxic compounds in liver by conjugating them with GSH, an intracellular thiol antioxidant (Vijayakumar *et.al.*, 2010). Many studies observed that vitamin C reducing plasma lipid peroxide levels, increasing GSH and enhance enzymic antioxidants (Osman *et.al*, 2010). Significant recovery in the level of antioxidant enzymes in diabetic rats administered with *Mollugo nudicaulis* may be due to correction in plasma insulin through beta cells stimulating effect of phyto ingredients present in it.

Earlier studies indicated that glibenclamide possess direct mechanism to enhance the level of antioxidant enzymes besides reducing the lipid peroxidation in diabetic animals (Rabbani *et.al.*, 2010). In present study also, glibenclamide significantly increased the level of antioxidant enzymes and decreased the lipid peroxidation by its antioxidant activity.

# CONCLUSION

The study showed that the anti-diabetic activity of the ethanolic extract of *Mollugo nudicaulis* against alloxan induced diabetic rats. Pharmacological studies are required to evaluate the exact mechanism of action and components present in it. Further work is in progress for isolation and identification of the components from *Mollugo nudicaulis*.

#### ACKNOWLEDGEMENTS

We, the authors are thankful to our Chancellor, Advisor, Vice Chancellor and Registrar of Karpagam University for providing facilities and encouragement.

International Journal of Applied Biology and Pharmaceutical Technology Page:517 Available online at <u>www.ijabpt.com</u>



# REFERENCES

L.R.Atmakuri and S.Dathi (2010). Current trends in herbal medicines. JPR. 3:109-113.

J.Brod and J.H.Sirota (1948). The renal clearance of endogenous creatinine in man. J Clin Invest, 27:645-54.

W.I.Caraway (1963) Uric acid In. Standard Methods of Clinical Chemistry. D.Seligson editors. Academic Press, New York, 4:239-47.

P.K.Chauhan, I.P.Pandey, V.K.Dhatwalia and V.Singh (2010). Anti-diabetic activity of etanolic and methanolic leaves extract of *Centella asiatica* on alloxan induced diabetic rats. IJPBS, 2:1-6.

V.Chitra, P.V.K.R.C.H.H.Varma, M.V.R.K.Raju, and K.J.Prakash (2010). Study of antidiabetic and free radical scavenging activity of the seed extract of *Strychnos nuxvomica*. Int J Pharm Pharm Sci, 2:106-110.

R.Dheer and P.Bhatnagar (2010). A study of the antidiabetic activity of *Barleria prionitis* Linn. Indian J Pharmacol, 42:70-73.

V.Gupta and M.Sharma (2010). Protective effect of *Cinnamomum tejpata* on lipid peroxide formation in isolated rat liver homogenate. Curr Res J Biol Sci, 2:246-249.

W.H.Habig, M.J.Pabst and W.B.Jakoby (1974). Glutathione S-transferase. The first enzymatic step in mercapturic acid formation. J Bio Chem, 249:7130-7139.

J.Hogberg, R.E.Larson, A.Kristoferson and S.Orrhenius (1974). NADPH-dependent reductase solubilized from microsomes by peroxidation and its activity. Biochem Biophys Res Commun, 56:836-842.

S.Kaimal, K.S.Sujatha and S.George (2010). Hypolipidaemic and antioxidant effects of fruits of *Musa AAA* (Chenkadali) in alloxan induced diabetic rats. Indian J Exp Biol, 48:165-173.

J.Kakadiya, M.Shah, and N.J.Shah (2010). Effect of nobivolol on serum diabetic marker and lipid profile in normal and streptozotocin-nicotinamide induced diabetic rats. RJPBCS, 1:329-334.

J.Kakadiya, M.Shah and N.Shah (2010). Glimepiride reduces on experimentally induced is chemical reperfusion in diabetic rats. IJABPT, 1:276-285.

E.J.King (1965). In "Practical Clinical Enzymology". H.J.Princeton editor. Van Nostrand Reinhold Co. Ltd, London, pp.83-93.

O.H.Lowry, N.J.Rosebrough, A.L.Farr, and R.J.Randall (1951). Protein measurement with the Folin Phenol reagent. J Biol Chem. 193:265-275.

H.Lueck (1965). In: Methods of Enzymatic Analysis. London: Academic Press.

N.Malviya, S.Jain and S.Malviya (2010). Antidiabetic potential of medicinal plants. APPHA. 67:113-118.

H.P.Misra and I.Fridovich (1972). The role of Superoxide anion in the antioxidant of epinephrine and a single assay of Superoxide dismutase. J Biol Chem, 247:3170-3175.

M.S.Moran, J.W.Depierre and B.Mannervik (1979). Levels of glutathione, glutathione reductase and glutathione-S-transferase activities in rat lung and liver. Biochim Biophys Acta, 582:67-78.

M.Muthulingam (2010). Antidiabetic efficacy of leaf extracts of *Asteracantha longifolia* (Linn.) Nees. on alloxan induced diabetics in male albino wistar rats. Int J Pharm Biomed Res, 1:28-34.

K.S.Nagesh (2008). Wound healing activity of *Mollugo nudicaulis* Lam: Field grown leaf versus *in vitro*derived Calli extracts on rats. Third congress of the World Union of wound healing societies. June 4-8, Toronto, Canada.

S.Natelson, M.L.Scott, and C.Beffa (1951). A rapid method for the estimation of urea in biological fluids. Am J Clin Pathol, 21:275-81.

C.O.Okoli, A.F.Ibiam, A.C.Ezike, P.A.Akah and T.C.Okoye (2010). Evaluation of antidiabetic potentials of *Phyllanthus niruri* in alloxan diabetic rats. Afr J Biotechnol, 9:248-259.

S.T.Omaye, J.D.Turabull, and H.E.Sauberlich (1979). Selected methods for the determination of ascorbic acid in animal cells, tissues and fluids. Method Enzymol. 62:1-11.

M.Osman, S.A.Fayed, G.I.Mohmoud and R.M.Romeilah (2010). Protective effects of Chitosan, ascorbic acid and *Gymnema Sylvestre* against hypercholesterolemia in male rats. Aust J Basic & Appl Sci, 4:89-98.

V.K.Sharma, S.Kumar, H.J.Patel and S.Hugar (2010). Hypoglycemic activity of *Ficus glomerata* in alloxan induced diabetic rats. IJPSRR, 1:18-22.

International Journal of Applied Biology and Pharmaceutical Technology Page:518 Available online at <u>www.ijabpt.com</u>



S.I.Rabbani, K.Devi and S.Khanam (2010). Protective role of glibenclamide against nicotinamidestreptozotocin induced nuclear damage in diabetic wistar rats. J Pharmacol Parmacother, 1:18-23.

S.Ragupathy, N.G.Steven, M.Maruthakkutti, B.Velusamy and M.M.UI-Huda (2008). Consensus of the 'Malasars' traditional aboriginal knowledge of medicinal plants in the Velliangiri holy hills, India. J Ethnobiol Ethnomed, 4:1-14.

N.R.Rathod, I.Raghuveer, H.R.Chitme and R.Chandra (2009). Free radical scavenging activity of *Calotropis gigantea* on streptozotocin-induced diabetic rats. Indian J Pharm Sci, 71:615-621.

I.K.Rao (2009). Structural study of Nano powder *Mollugo nudicaulis* medical herb. International conference on materials for advanced technologies and International union of materials research societies-International conference in Asia. June 28-July 3, Singapore.

J.T.Rotruck, A.L.Pope and H.S.Ganther (1973). Selenium: Biochemical role as a component of glutathione peroxidase purification and assay. Science, 179:588-590.

M.S.Sikarwar, M.B.Patil (2010). Antidiabetic activity of *Crateva nurvala* stem bark extracts in alloxaninduced diabetic rats. J Pharm Bioall Sci. 2:18-21.

A.U.Tatiya, U.V.Deore, P.G.Jain and S.J.Surana (2010). Hypoglycemic potential of *Bridelia retusa* bark in albino rats. Asian J Biol Sci, 4:84-89.

M.Vijayakumar, A.Balasubramaniam, R.Manivannan and N.S.Kumar (2010). Antioxidant potential of ethanolic extract of *Bauhinia tomentosa* (Linn) flower. RJPBCS, 1:143-147.

\*\*\*\*\*

International Journal of Applied Biology and Pharmaceutical Technology Page:519 Available online at <u>www.ijabpt.com</u>